

VIII. IRODALOMJEGYZÉK

1. Moskowitz, M.A., *The neurobiology of vascular head pain*. Ann Neurol, 1984. **16**(2): p. 157-68.
2. Buzzi, M.G. and M.A. Moskowitz, *The antimigraine drug, sumatriptan (GR43175), selectively blocks neurogenic plasma extravasation from blood vessels in dura mater*. Br J Pharmacol, 1990. **99**(1): p. 202-6.
3. Saito, K., S. Markowitz, and M.A. Moskowitz, *Ergot alkaloids block neurogenic extravasation in dura mater: proposed action in vascular headaches*. Ann Neurol, 1988. **24**(6): p. 732-7.
4. Tajti, J., et al., *Messenger molecules and receptor mRNA in the human trigeminal ganglion*. J Auton Nerv Syst, 1999. **76**(2-3): p. 176-83.
5. Gulbenkian, S., R. Uddman, and L. Edvinsson, *Neuronal messengers in the human cerebral circulation*. Peptides, 2001. **22**(6): p. 995-1007.
6. Lennerz, J.K., et al., *Calcitonin receptor-like receptor (CLR), receptor activity-modifying protein 1 (RAMP1), and calcitonin gene-related peptide (CGRP) immunoreactivity in the rat trigeminovascular system: differences between peripheral and central CGRP receptor distribution*. J Comp Neurol, 2008. **507**(3): p. 1277-99.
7. Goadsby, P.J., L. Edvinsson, and R. Ekman, *Release of vasoactive peptides in the extracerebral circulation of humans and the cat during activation of the trigeminovascular system*. Ann Neurol, 1988. **23**(2): p. 193-6.
8. Goadsby, P.J., L. Edvinsson, and R. Ekman, *Vasoactive peptide release in the extracerebral circulation of humans during migraine headache*. Ann Neurol, 1990. **28**(2): p. 183-7.
9. Nicolodi, M. and E. Del Bianco, *Sensory neuropeptides (substance P, calcitonin gene-related peptide) and vasoactive intestinal polypeptide in human saliva: their pattern in migraine and cluster headache*. Cephalalgia, 1990. **10**(1): p. 39-50.
10. Tuka, B., et al., *Alterations in PACAP-38-like immunoreactivity in the plasma during ictal and interictal periods of migraine patients*. Cephalalgia, 2013.
11. Lassen, L.H., et al., *CGRP may play a causative role in migraine*. Cephalalgia, 2002. **22**(1): p. 54-61.
12. Uddman, R., et al., *Innervation of the feline cerebral vasculature by nerve fibers containing calcitonin gene-related peptide: trigeminal origin and co-existence with substance P*. Neurosci Lett, 1985. **62**(1): p. 131-6.

13. Edvinsson, L., et al., *Neurokinin A in cerebral vessels: characterization, localization and effects in vitro*. Regul Pept, 1988. **20**(3): p. 181-97.
14. Edvinsson, L., H. Hara, and R. Uddman, *Retrograde tracing of nerve fibers to the rat middle cerebral artery with true blue: colocalization with different peptides*. J Cereb Blood Flow Metab, 1989. **9**(2): p. 212-8.
15. Edvinsson, L., et al., *Demonstration of neuropeptide containing nerves and vasomotor responses to perivascular peptides in human cerebral arteries*. Cephalalgia, 1994. **14**(2): p. 88-96.
16. Suzuki, N., J.E. Hardebo, and C. Owman, *Origins and pathways of cerebrovascular nerves storing substance P and calcitonin gene-related peptide in rat*. Neuroscience, 1989. **31**(2): p. 427-38.
17. Cunha e Sa, M., et al., *Innervation pattern of malformative cortical vessels in Sturge-Weber disease: an histochemical, immunohistochemical, and ultrastructural study*. Neurosurgery, 1997. **41**(4): p. 872-6; discussion 876-7.
18. Edvinsson, L., et al., *Innervation of the human middle meningeal artery: immunohistochemistry, ultrastructure, and role of endothelium for vasomotility*. Peptides, 1998. **19**(7): p. 1213-25.
19. Gulbenkian, S., et al., *Ultrastructural evidence for the coexistence of calcitonin gene-related peptide and substance P in secretory vesicles of peripheral nerves in the guinea pig*. J Neurocytol, 1986. **15**(4): p. 535-42.
20. Tajti, J., et al., *Migraine is a neuronal disease*. J Neural Transm, 2011. **118**(4): p. 511-24.
21. Samsam, M., et al., *Alterations in neurokinin A-, substance P- and calcitonin gene-related peptide immunoreactivities in the caudal trigeminal nucleus of the rat following electrical stimulation of the trigeminal ganglion*. Neurosci Lett, 1999. **261**(3): p. 179-82.
22. Miyata, A., et al., *Isolation of a novel 38 residue-hypothalamic polypeptide which stimulates adenylate cyclase in pituitary cells*. Biochem Biophys Res Commun, 1989. **164**(1): p. 567-74.
23. Arimura, A., et al., *Tissue distribution of PACAP as determined by RIA: highly abundant in the rat brain and testes*. Endocrinology, 1991. **129**(5): p. 2787-9.
24. Ghatei, M.A., et al., *Distribution, molecular characterization of pituitary adenylate cyclase-activating polypeptide and its precursor encoding messenger RNA in human and rat tissues*. J Endocrinol, 1993. **136**(1): p. 159-66.

25. Hannibal, J., et al., *Gene expression of pituitary adenylate cyclase activating polypeptide (PACAP) in the rat hypothalamus*. Regul Pept, 1995. **55**(2): p. 133-48.
26. Hannibal, J., et al., *Pituitary adenylate cyclase-activating peptide gene expression in corticotropin-releasing factor-containing parvicellular neurons of the rat hypothalamic paraventricular nucleus is induced by colchicine, but not by adrenalectomy, acute osmotic, ether, or restraint stress*. Endocrinology, 1995. **136**(9): p. 4116-24.
27. Arimura, A. and S. Shioda, *Pituitary adenylate cyclase activating polypeptide (PACAP) and its receptors: neuroendocrine and endocrine interaction*. Front Neuroendocrinol, 1995. **16**(1): p. 53-88.
28. Rawlings, S.R. and M. Hezareh, *Pituitary adenylate cyclase-activating polypeptide (PACAP) and PACAP/vasoactive intestinal polypeptide receptors: actions on the anterior pituitary gland*. Endocr Rev, 1996. **17**(1): p. 4-29.
29. Mikkelsen, J.D., et al., *Pituitary adenylate cyclase activating peptide-38 (PACAP-38), PACAP-27, and PACAP related peptide (PRP) in the rat median eminence and pituitary*. J Neuroendocrinol, 1995. **7**(1): p. 47-55.
30. Filipsson, K., et al., *PACAP stimulates insulin secretion but inhibits insulin sensitivity in mice*. Am J Physiol, 1998. **274**(5 Pt 1): p. E834-42.
31. Watanabe, T., et al., *PACAP stimulates catecholamine release from adrenal medulla: a novel noncholinergic secretagogue*. Am J Physiol, 1995. **269**(5 Pt 1): p. E903-9.
32. Steenstrup, B.R., et al., *Pituitary adenylate cyclase-activating polypeptide: occurrence and relaxant effect in female genital tract*. Am J Physiol, 1995. **269**(1 Pt 1): p. E108-17.
33. Cardell, L.O., et al., *Pituitary adenylate cyclase activating peptide (PACAP) in guinea-pig lung: distribution and dilatory effects*. Regul Pept, 1991. **36**(3): p. 379-90.
34. Hauser-Kronberger, C., et al., *Distribution of two VIP-related peptides, helospectin and pituitary adenylate cyclase activating peptide (PACAP), in the human upper respiratory system*. Regul Pept, 1996. **65**(3): p. 203-9.
35. Hauser-Kronberger, C., et al., *Neuropeptides in human salivary (submandibular and parotid) glands*. Acta Otolaryngol, 1992. **112**(2): p. 343-8.
36. Vincze, E., et al., *Pituitary adenylate cyclase activating polypeptide (PACAP) is present in human and cat gastric glands*. Peptides, 1999. **20**(8): p. 937-41.

37. Mao, Y.K., et al., *Locations and molecular forms of PACAP and sites and characteristics of PACAP receptors in canine ileum*. Am J Physiol, 1998. **274**(1 Pt 1): p. G217-25.
38. Edvinsson, L. and R. Uddman, *Neurobiology in primary headaches*. Brain Res Brain Res Rev, 2005. **48**(3): p. 438-56.
39. Christiansen, T., et al., *Immunoreactivity of NOS, CGRP, PACAP, SP and VIP in the trigeminal nucleus caudalis and in the cervical spinal cord C1 and C2 of the cat*. J Headache Pain, 2003. **4**: p. 156–163.
40. Takeuchi, K., et al., *Stimulatory effect of PACAP on gastroduodenal bicarbonate secretion in rats*. J Physiol Paris, 1997. **91**(3-5): p. 229-34.
41. Mungan, Z., et al., *Effect of pituitary adenylate cyclase activating polypeptide on rat pancreatic exocrine secretion*. Peptides, 1991. **12**(3): p. 559-62.
42. Buscail, L., et al., *Stimulation of rat pancreatic tumoral AR4-2J cell proliferation by pituitary adenylate cyclase-activating peptide*. Gastroenterology, 1992. **103**(3): p. 1002-8.
43. Douziech, N., et al., *Growth effects of regulatory peptides and intracellular signaling routes in human pancreatic cancer cell lines*. Endocrine, 1998. **9**(2): p. 171-83.
44. Bhawe, S.V. and P.L. Hoffman, *Phosphatidylinositol 3'-OH kinase and protein kinase A pathways mediate the anti-apoptotic effect of pituitary adenylate cyclase-activating polypeptide in cultured cerebellar granule neurons: modulation by ethanol*. J Neurochem, 2004. **88**(2): p. 359-69.
45. Falluel-Morel, A., et al., *Pituitary adenylate cyclase-activating polypeptide prevents the effects of ceramides on migration, neurite outgrowth, and cytoskeleton remodeling*. Proc Natl Acad Sci U S A, 2005. **102**(7): p. 2637-42.
46. Vallejo, I. and M. Vallejo, *Pituitary adenylate cyclase-activating polypeptide induces astrocyte differentiation of precursor cells from developing cerebral cortex*. Mol Cell Neurosci, 2002. **21**(4): p. 671-83.
47. Mabuchi, T., et al., *Pituitary adenylate cyclase-activating polypeptide is required for the development of spinal sensitization and induction of neuropathic pain*. J Neurosci, 2004. **24**(33): p. 7283-91.
48. Huang, M., H. Shirahase, and O.P. Rorstad, *Comparative study of vascular relaxation and receptor binding by PACAP and VIP*. Peptides, 1993. **14**(4): p. 755-62.
49. Tong, S., et al., *Pituitary adenylate cyclase-activating polypeptide dilates cerebral arterioles of newborn pigs*. Proc Soc Exp Biol Med, 1993. **203**(3): p. 343-7.

50. Warren, J.B., et al., *Pituitary adenylate cyclase-activating polypeptide: a novel, long-lasting, endothelium-independent vasorelaxant*. Eur J Pharmacol, 1991. **197**(2-3): p. 131-4.
51. Tuka, B., et al., *Peripheral and central alterations of pituitary adenylate cyclase activating polypeptide-like immunoreactivity in the rat in response to activation of the trigeminovascular system*. Peptides, 2012. **33**(2): p. 307-16.
52. Schytz, H.W., et al., *PACAP38 induces migraine-like attacks in patients with migraine without aura*. Brain, 2009. **132**(Pt 1): p. 16-25.
53. Kitazono, T., et al., *Role of potassium channels in cerebral blood vessels*. Stroke, 1995. **26**(9): p. 1713-23.
54. Henning, R.J. and D.R. Sawmiller, *Vasoactive intestinal peptide: cardiovascular effects*. Cardiovasc Res, 2001. **49**(1): p. 27-37.
55. Kulka, M., et al., *Neuropeptides activate human mast cell degranulation and chemokine production*. Immunology, 2008. **123**(3): p. 398-410.
56. Theoharides, T.C., et al., *The role of mast cells in migraine pathophysiology*. Brain Res Brain Res Rev, 2005. **49**(1): p. 65-76.
57. Gilfillan, A.M. and C. Tkaczyk, *Integrated signalling pathways for mast-cell activation*. Nat Rev Immunol, 2006. **6**(3): p. 218-30.
58. Lassen, L.H., et al., *Nitric oxide synthase inhibition in migraine*. Lancet, 1997. **349**(9049): p. 401-2.
59. Sicuteri, F., et al., *Unmasking latent dysnociception in healthy subjects*. Headache, 1987. **27**(4): p. 180-5.
60. Hsi, D.H., et al., *Headache response to glyceryl trinitrate in patients with and without obstructive coronary artery disease*. Heart, 2005. **91**(9): p. 1164-6.
61. Pardutz, A., et al., *Systemic nitroglycerin increases nNOS levels in rat trigeminal nucleus caudalis*. Neuroreport, 2000. **11**(14): p. 3071-5.
62. Pardutz, A., et al., *Oestrogen-modulated increase of calmodulin-dependent protein kinase II (CamKII) in rat spinal trigeminal nucleus after systemic nitroglycerin*. Cephalalgia, 2007. **27**(1): p. 46-53.
63. Tassorelli, C., et al., *Parthenolide is the component of tanacetum parthenium that inhibits nitroglycerin-induced Fos activation: studies in an animal model of migraine*. Cephalalgia, 2005. **25**(8): p. 612-21.

64. Centini, C. and O. Pompeiano, *Sleep research in space: expression of immediate early genes in forebrain structures of rats during the nasa neurolab mission (STS-90)*. Arch Ital Biol, 2007. **145**(2): p. 117-50.
65. Tallaksen-Greene, S.J., et al., *Excitatory amino acid binding sites in the trigeminal principal sensory and spinal trigeminal nuclei of the rat*. Neurosci Lett, 1992. **141**(1): p. 79-83.
66. Vikelis, M. and D.D. Mitsikostas, *The role of glutamate and its receptors in migraine*. CNS Neurol Disord Drug Targets, 2007. **6**(4): p. 251-7.
67. Lauritzen, M., *Pathophysiology of the migraine aura. The spreading depression theory*. Brain, 1994. **117** (Pt 1): p. 199-210.
68. Goadsby, P.J. and J.D. Classey, *Glutamatergic transmission in the trigeminal nucleus assessed with local blood flow*. Brain Res, 2000. **875**(1-2): p. 119-24.
69. Sugihara, H., et al., *Structures and properties of seven isoforms of the NMDA receptor generated by alternative splicing*. Biochem Biophys Res Commun, 1992. **185**(3): p. 826-32.
70. Monyer, H., et al., *Heteromeric NMDA receptors: molecular and functional distinction of subtypes*. Science, 1992. **256**(5060): p. 1217-21.
71. Meguro, H., et al., *Functional characterization of a heteromeric NMDA receptor channel expressed from cloned cDNAs*. Nature, 1992. **357**(6373): p. 70-4.
72. Mitsikostas, D.D., et al., *The NMDA receptor antagonist MK-801 reduces capsaicin-induced c-fos expression within rat trigeminal nucleus caudalis*. Pain, 1998. **76**(1-2): p. 239-48.
73. Mitsikostas, D.D., et al., *Non-NMDA glutamate receptors modulate capsaicin induced c-fos expression within trigeminal nucleus caudalis*. Br J Pharmacol, 1999. **127**(3): p. 623-30.
74. Storer, R.J. and P.J. Goadsby, *Trigeminovascular nociceptive transmission involves N-methyl-D-aspartate and non-N-methyl-D-aspartate glutamate receptors*. Neuroscience, 1999. **90**(4): p. 1371-6.
75. Filla, S.A., et al., *Ethyl (3S,4aR,6S,8aR)-6-(4-ethoxycarbonylimidazol-1-ylmethyl)decahydroiso-quinoline-3-carboxylic ester: a prodrug of a GluR5 kainate receptor antagonist active in two animal models of acute migraine*. J Med Chem, 2002. **45**(20): p. 4383-6.
76. Kolhekar, R., S. Murphy, and G.F. Gebhart, *Thalamic NMDA receptors modulate inflammation-produced hyperalgesia in the rat*. Pain, 1997. **71**(1): p. 31-40.

77. Classey, J.D., Y.E. Knight, and P.J. Goadsby, *The NMDA receptor antagonist MK-801 reduces Fos-like immunoreactivity within the trigeminocervical complex following superior sagittal sinus stimulation in the cat*. Brain Res, 2001. **907**(1-2): p. 117-24.
78. Haley, J.E., A.F. Sullivan, and A.H. Dickenson, *Evidence for spinal N-methyl-D-aspartate receptor involvement in prolonged chemical nociception in the rat*. Brain Res, 1990. **518**(1-2): p. 218-26.
79. Woolf, C.J. and S.W. Thompson, *The induction and maintenance of central sensitization is dependent on N-methyl-D-aspartic acid receptor activation; implications for the treatment of post-injury pain hypersensitivity states*. Pain, 1991. **44**(3): p. 293-9.
80. Domoki, F., et al., *N-methyl-D-aspartate-induced vasodilation is mediated by endothelium-independent nitric oxide release in piglets*. Am J Physiol Heart Circ Physiol, 2002. **282**(4): p. H1404-9.
81. Ayata, C. and M.A. Moskowitz, *Cortical spreading depression confounds concentration-dependent pial arteriolar dilation during N-methyl-D-aspartate superfusion*. Am J Physiol Heart Circ Physiol, 2006. **290**(5): p. H1837-41.
82. Kaube, H., et al., *Aura in some patients with familial hemiplegic migraine can be stopped by intranasal ketamine*. Neurology, 2000. **55**(1): p. 139-41.
83. Rajda, C., et al., *Amino acids in the saliva of patients with migraine*. Headache, 1999. **39**(9): p. 644-9.
84. Jhamandas, K., et al., *Quinolate-induced cortical cholinergic damage: modulation by tryptophan metabolites*. Brain Res, 1990. **529**(1-2): p. 185-91.
85. de Carvalho, L.P., P. Bochet, and J. Rossier, *The endogenous agonist quinolinic acid and the non endogenous homoquinolinic acid discriminate between NMDAR2 receptor subunits*. Neurochem Int, 1996. **28**(4): p. 445-52.
86. Rios, C. and A. Santamaria, *Quinolinic acid is a potent lipid peroxidant in rat brain homogenates*. Neurochem Res, 1991. **16**(10): p. 1139-43.
87. Behan, W.M., et al., *Oxidative stress as a mechanism for quinolinic acid-induced hippocampal damage: protection by melatonin and deprenyl*. Br J Pharmacol, 1999. **128**(8): p. 1754-60.
88. Stone, T.W., *Kynurenines in the CNS: from endogenous obscurity to therapeutic importance*. Prog Neurobiol, 2001. **64**(2): p. 185-218.

89. Stone, T.W., *Endogenous neurotoxins from tryptophan*. Toxicol, 2001. **39**(1): p. 61-73.
90. Smith, D.H., et al., *Effects of the excitatory amino acid receptor antagonists kynurenate and indole-2-carboxylic acid on behavioral and neurochemical outcome following experimental brain injury*. J Neurosci, 1993. **13**(12): p. 5383-92.
91. Gigler, G., et al., *Neuroprotective effect of L-kynurenine sulfate administered before focal cerebral ischemia in mice and global cerebral ischemia in gerbils*. Eur J Pharmacol, 2007. **564**(1-3): p. 116-22.
92. Nasstrom, J., U. Karlsson, and C. Post, *Antinociceptive actions of different classes of excitatory amino acid receptor antagonists in mice*. Eur J Pharmacol, 1992. **212**(1): p. 21-9.
93. Hajos, M. and G. Engberg, *Kynurenic acid blocks chemogenic nociception*. J Pharm Pharmacol, 1990. **42**(5): p. 373-4.
94. Song, X.J. and Z.Q. Zhao, *Differential effects of NMDA and non-NMDA receptor antagonists on spinal cutaneous vs muscular nociception in the cat*. Neuroreport, 1993. **4**(1): p. 17-20.
95. Knyihar-Csillik, E., et al., *Kynurenine aminotransferase in the supratentorial dura mater of the rat: effect of stimulation of the trigeminal ganglion*. Exp Neurol, 2004. **186**(2): p. 242-7.
96. Birch, P.J., C.J. Grossman, and A.G. Hayes, *Kynurenate and FG9041 have both competitive and non-competitive antagonist actions at excitatory amino acid receptors*. Eur J Pharmacol, 1988. **151**(2): p. 313-5.
97. Hilmas, C., et al., *The brain metabolite kynurenic acid inhibits alpha7 nicotinic receptor activity and increases non-alpha7 nicotinic receptor expression: physiopathological implications*. J Neurosci, 2001. **21**(19): p. 7463-73.
98. Vecsei, L., et al., *Kynurenine and probenecid inhibit pentylenetetrazol- and NMDLA-induced seizures and increase kynurenic acid concentrations in the brain*. Brain Res Bull, 1992. **28**(2): p. 233-8.
99. Rozsa, E., et al., *The pentylenetetrazole-induced activity in the hippocampus can be inhibited by the conversion of L-kynurenine to kynurenic acid: an in vitro study*. Brain Res Bull, 2008. **76**(5): p. 474-9.
100. Knyihar-Csillik, E., et al., *Kynurenine in combination with probenecid mitigates the stimulation-induced increase of c-fos immunoreactivity of the rat caudal trigeminal*

- nucleus in an experimental migraine model. J Neural Transm*, 2007. **114**(4): p. 417-21.
101. Knyihar-Csillik, E., et al., *The kynurenate analog SZR-72 prevents the nitroglycerol-induced increase of c-fos immunoreactivity in the rat caudal trigeminal nucleus: comparative studies of the effects of SZR-72 and kynurenic acid. Neurosci Res*, 2008. **61**(4): p. 429-32.
 102. Vamos, E., et al., *l-kynurenine combined with probenecid and the novel synthetic kynurenic acid derivative attenuate nitroglycerin-induced nNOS in the rat caudal trigeminal nucleus. Neuropharmacology*, 2009. **57**(4): p. 425-9.
 103. Vamos, E., et al., *Kynurenate derivative attenuates the nitroglycerin-induced CamKIIalpha and CGRP expression changes. Headache*, 2010. **50**(5): p. 834-43.
 104. Harrington, M.E., et al., *Pituitary adenylate cyclase activating peptide phase shifts circadian rhythms in a manner similar to light. J Neurosci*, 1999. **19**(15): p. 6637-42.
 105. Ohsawa, M., et al., *Modulation of nociceptive transmission by pituitary adenylate cyclase activating polypeptide in the spinal cord of the mouse. Pain*, 2002. **100**(1-2): p. 27-34.
 106. MacDonald, J.F., M.F. Jackson, and M.A. Beazely, *G protein-coupled receptors control NMDARs and metaplasticity in the hippocampus. Biochim Biophys Acta*, 2007. **1768**(4): p. 941-51.
 107. Endo, K., et al., *Neuroprotective effect of PACAP against NMDA-induced retinal damage in the mouse. J Mol Neurosci*, 2011. **43**(1): p. 22-9.
 108. Fukuchi, M., A. Tabuchi, and M. Tsuda, *Activity-dependent transcriptional activation and mRNA stabilization for cumulative expression of pituitary adenylate cyclase-activating polypeptide mRNA controlled by calcium and cAMP signals in neurons. J Biol Chem*, 2004. **279**(46): p. 47856-65.
 109. Tabuchi, A., *Synaptic plasticity-regulated gene expression: a key event in the long-lasting changes of neuronal function. Biol Pharm Bull*, 2008. **31**(3): p. 327-35.
 110. Fanciullacci, M., M. Alessandri, and A. Del Rosso, *Dopamine involvement in the migraine attack. Funct Neurol*, 2000. **15 Suppl 3**: p. 171-81.
 111. Cherchi, A., et al., *Role of dopaminergic system in migraine. J Headache Pain* 2001. **2**(Suppl 1): p. 47-49.
 112. Peterfreund, R.A., B.E. Kosofsky, and J.S. Fink, *Cellular localization of dopamine D2 receptor messenger RNA in the rat trigeminal ganglion. Anesth Analg*, 1995. **81**(6): p. 1181-5.

113. Tomita, H., et al., *Inducible cAMP early repressor (ICER) is a negative-feedback regulator of cardiac hypertrophy and an important mediator of cardiac myocyte apoptosis in response to beta-adrenergic receptor stimulation*. Circ Res, 2003. **93**(1): p. 12-22.
114. Vela, J., A. Akopian, and K. Hargreaves, *Modulation of peripheral trigeminal nociceptors by beta adrenergic receptor-1*. The Journal of Pain, 2006. **7**(4): p. S23.
115. D'Andrea, G., et al., *The role of tyrosine metabolism in the pathogenesis of chronic migraine*. Cephalalgia, 2013.
116. Silberstein, S.D., *Preventive migraine treatment*. Neurol Clin, 2009. **27**(2): p. 429-43.
117. Neri, G., et al., *Pituitary adenylate-cyclase activating peptide enhances aldosterone secretion of human adrenal gland: evidence for an indirect mechanism, probably involving the local release of catecholamines*. J Clin Endocrinol Metab, 1996. **81**(1): p. 169-73.
118. Breault, L., et al., *Occurrence and effect of PACAP in the human fetal adrenal gland*. Ann N Y Acad Sci, 2000. **921**: p. 429-33.

IX. KÖSZÖNETNYILVÁNÍTÁS

Ezúton szeretnék köszönetet mondani Dr. Vécsei László tanszékvezető professzor úrnak, az SZTE-ÁOK Neurológiai Klinika igazgatójának, hogy helyet biztosított gyakorlatszerzésemre.

Köszönet szeretnék mondani témavezetőmnek Dr. Tajti János egyetemi docensnek, hogy lehetőséget adott arra, hogy részt vegyek az aktuális kísérletekben és szakmai észrevételeivel valamint hasznos útmutatásaival hozzájárult a kutatómunkámhoz és dolgozatom elkészüléséhez.

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Nem utolsó sorban hatalmas köszönet illeti belső konzulensemét, Dr. Fekete Éva professzorasszonyt, aki segítőkészségével irányította ifjú kutatói munkásságomat.

X. TÉTELES SZAKMAI HOZZÁJÁRULÁS

Alulírott Mészáros Ádám igazolom, hogy a dolgozatomban leírt munkafolyamatok közül:

- az állatok trigeminalis rendszerének elektromos ingerlését,
- transcárdialis perfúzióját,
- és a minták preparálását

végeztem el, illetve részt vettem a statisztikai elemzésben.



Mészáros Ádám

Szeged, 2014. november 12.